

Research Article

Mortality Risk of Juvenile Japanese Sea Cucumber *Apostichopus japonicus* by the Sympatrically Occurring Hermit Crab *Paguristes ortmanni*

Nobuharu Inaba ^{1,2}, Takuma Matsumoto,¹ Yuji Anaguchi,³ and Kohei Matsuno^{4,5}

¹Civil Engineering Research Institute for Cold Region, Public Works Research Institute, Sapporo 062-8602, Hokkaido, Japan

²School of Biological, Earth and Environmental Sciences, Faculty of Science, University of New South Wales, Sydney 2052, New South Wales, Australia

³Ocean Construction Co. Ltd., Kurashiki 711-0924, Okayama, Japan

⁴Faculty/Graduate School of Fisheries Sciences, Hokkaido University, Hakodate 041-8611, Hokkaido, Japan

⁵Arctic Research Center, Hokkaido University, Sapporo 001-0021, Hokkaido, Japan

Correspondence should be addressed to Nobuharu Inaba; inaba-n@ceri.go.jp

Received 15 October 2024; Accepted 22 February 2025

Copyright © 2025 Nobuharu Inaba et al. Aquaculture Research published by John Wiley & Sons Ltd. This is an open access article under the terms of the Creative Commons Attribution License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

The density of the hermit crab, *Paguristes ortmanni*, in the artificial reef of juvenile *Apostichopus japonicus* increased almost threefold from June to December 2018. Calcareous ossicles of *A. japonicus* were identified from the stomach contents in 28.8% of the hermit crabs (18.6% in males and 10.2% in females) in December; season hatchery-produced juveniles are frequently introduced into the wild in Hokkaido, Japan. The mortality rate of juvenile *A. japonicus* by *P. ortmanni* was estimated to be 2.5 ± 2.4 individuals day⁻¹ based on laboratory predation experiments. Interestingly, 3% to 5% of individuals survived despite being attacked and injured in all trials, escaping on the shells of hermit crabs. Over 50% of females in the ossicle-not-detected group had shield lengths (SLs) smaller than the smallest individual in the ossicle-detected group. The average SL of the ossicle-detected group in females was significantly higher ($p < 0.01$) than that of the not-detected group, indicating an increased predation risk for *A. japonicus* juveniles when larger female *P. ortmanni* were present. The present study offers new insights into the predatory behavior of *P. ortmanni* toward *A. japonicus* juveniles, showing that these sympatric hermit crabs present a considerably high mortality risk to *A. japonicus* juveniles. It also emphasizes the importance of implementing appropriate measures to protect juveniles from predators during the release process, providing an essential viewpoint for enhancing and rebuilding the wild population of commercially important endangered *A. japonicus*.

Keywords: *Apostichopus japonicus*; hermit crab; juvenile; *Paguristes*; predator; sea cucumber

1. Introduction

Sea cucumbers are marine invertebrates classified in the phylum Echinodermata, class Holothuroidea [1]. Sea cucumbers have been traditionally harvested in China for over a thousand years [2] and have been primarily consumed by East Asian populations, including Japan, for centuries [3]. China dominates the global sea cucumber trade, and the number of countries exporting sea cucumbers to China increased from 35 to 83 between 1996 and 2011 [4]. Concurrently, seven sea cucumber species traded in China are listed as “endangered”

on the International Union for Conservation of Nature (IUCN) Red List [5]. *Apostichopus japonicus* Selenka, 1867 (known as Japanese sea cucumber), a common temperate sea cucumber species inhabiting the coast of Russia, Korea, China, Japan, and the United States [6], is one of the seven endangered sea cucumber species [7]. Of these, *A. japonicus*, distributed in Hokkaido, the northernmost island of Japan, is characterized by its distinctive morphology, particularly the six rows of relatively sharp papillae on the dorsal and ventral surfaces [8]. The quantity and morphology of these papillae significantly

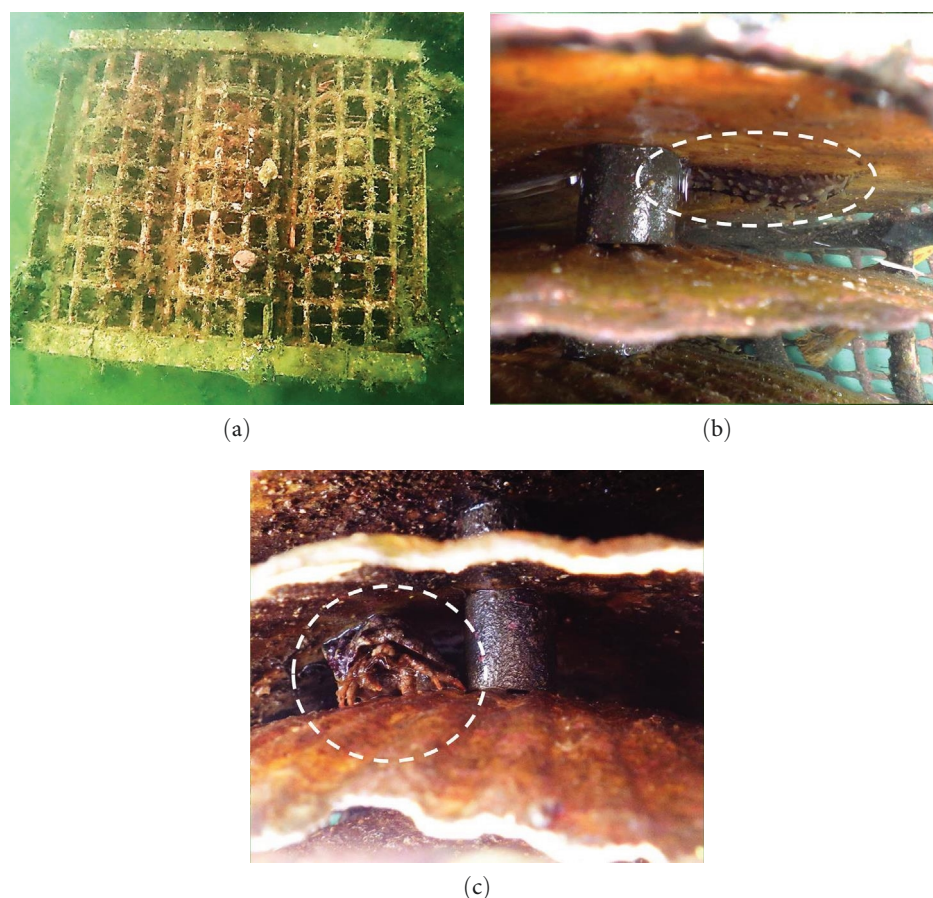


FIGURE 1: An underwater artificial intermediate reef for sea cucumber (a), a juvenile sea cucumber *A. japonicus* (b), and hermit crab *P. ortmanni* (c) observed inside the reef.

influence market value, with *A. japonicus* from Hokkaido commanding premium prices [8, 9].

Despite the early implementation of protective measures in Japan [10], wild populations of *A. japonicus* have been reported to have declined by 30% over the past 30 years [6]. Elevated fishing pressure and the emergence of systematic, large-scale poaching operations have been identified as critical concerns [3, 11], while habitat degradation due to coastal anthropogenic activities and numerous environmental stressors associated with climate change have been extensively discussed as factors contributing to the population decline of sea cucumbers [3, 12, 13]. Moreover, given that *A. japonicus* employs broadcast spawning as its mode of fertilization [14], insufficient numbers of matured males and females in close proximity during spawning events may significantly impair fertilization success, thereby posing a critical risk to population recovery [15]. In other words, once sea cucumber populations have declined below a certain threshold, in addition to the conventional protective measures, introducing hatchery-produced juveniles into natural habitats is the essential strategy for stock restoration [16]. In Japan, hatchery-produced *A. japonicus* juveniles have been widely released. For example, ~15 million juveniles (>5 mm) were released in Hokkaido in 2020 [17].

Previous studies have suggested that predators of adult sea cucumbers are relatively few [18, 19] due to their extensive

antipredatory behaviors, such as producing saponins as feeding deterrents [20], contracting body wall muscle [21, 22], and controlling buoyancy and strength of attachment when detached from a substratum [23, 24] and ejection of internal organs [25]. Meanwhile, predation is often observed as the most crucial factor of early juvenile mortality for a wide range of marine benthic invertebrate species [26–28]. Some studies have reported intensive predation on juvenile sea cucumbers, with *Holothuria scabra* being preyed upon by fish [29] and decapod crustaceans [30–32] and *A. japonicus* by the kelp crab [33]. These studies emphasize the importance of expanding our knowledge on sea cucumber predators and the impact of predation, particularly on the early juvenile stages of sea cucumbers.

In this study, we collected and measured the most dominant hermit crab, *Paguristes ortmanni* Miyake 1978, within *A. japonicus* artificial intermediate reefs composed of stacked scallop shells (Figures 1a–c), deployed at a fishing port in southwestern Hokkaido, Japan, between June and December 2018. The stomach contents of *P. ortmanni* were subjected to microscopic examination. Additionally, laboratory experiments were conducted to estimate the mortality rate of juvenile *A. japonicus* (green type) commonly distributed in the Hokkaido region [34] by *P. ortmanni*, and predator and prey behaviors were observed.

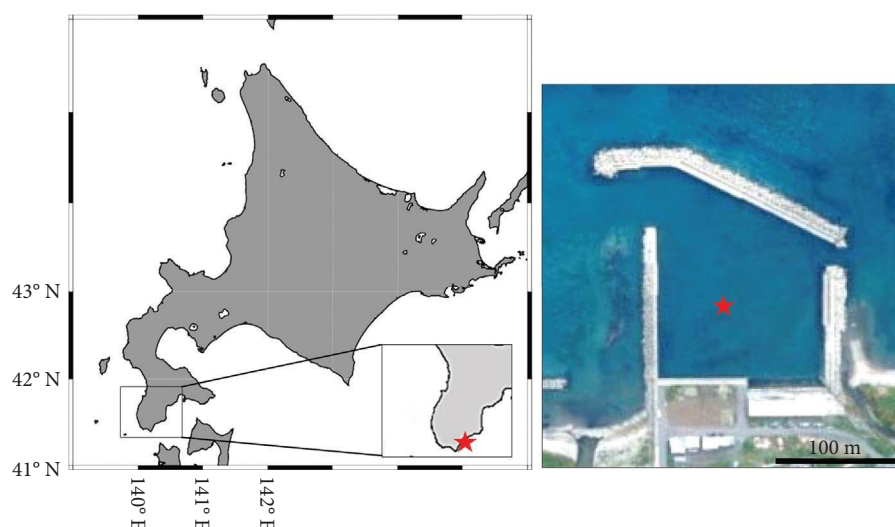


FIGURE 2: The fishing port where samplings were conducted, located in southwestern Hokkaido, Japan.

2. Material and Methods

2.1. Field Sampling, Measurement, and Observation of Stomach Contents. An artificial intermediate sea cucumber reef for *A. japonicus*, previously deployed at a depth of 4–5 m at a fishing port located in southwestern Hokkaido, Japan (41°27'44.8" N 140°14'52.7" E; Figure 2), was carefully wrapped in underwater nets and landed every 3 months from June 7 to December 6, 2018, to collect all hermit crabs inhabiting the inside of a reef. The collected hermit crabs were preserved using 70% ethanol immediately after sampling on-site and brought back to a laboratory, and *P. ortmanni* were identified [33, 35–37]. Body size measurements (shield length [SL] and wet weight) of all samples ($n = 97$) were performed using a digital vernier caliper to the nearest 0.01 mm (Figure 3a). Sex was determined from the gonopore position [38]. After these measurements, the stomach was dissected with a clean scalpel and tweezers to obtain food content from all of the *P. ortmanni* collected in December ($n = 59$), the season when hatchery-produced juveniles are often released in Hokkaido. The content was dissolved in a sodium hypochlorite solution (NaClO, 5%), and the dissolved content was microscopically observed using Axiovert 135 (Zeiss, Germany) to detect the ossicles of *A. japonicus* [33, 39]. The population density of *P. ortmanni* per unit volume was estimated based on the size of the artificial reef (0.5 m length \times 0.3 m width \times 0.15 m height). Measured values of SL and wet weight were tested using the Kruskal–Wallis test, followed by Dunn's multiple comparisons test to compare differences between samplings. The ossicle detection and non-detection rates between males and females were analyzed using the chi-square test. Using Welch's *t*-test, the differences in the body size index of the SL and the wet weight between the ossicle-detected and not-detected groups were also compared. The analysis was conducted after detecting outliers using the interquartile range (IQR) method and removing them from the dataset [40].

2.2. Estimation of Mortality Rate by Laboratory Experiments. Nine *P. ortmanni* (two males and seven females) collected at

the same sampling site described above on June 25, 2019, were gently placed into a cool, aerated plastic bucket and brought back to the laboratory alive. *P. ortmanni* were separately maintained in small aquaria (31 cm long \times 18 cm wide) at $13 \pm 1^\circ\text{C}$ based on the local water temperature measured in December 2018. A 12-h light/12-h dark cycle was applied, and the depth of the seawater in the aquaria was adjusted to 3 cm, the depth at which the studied *P. ortmanni* were confirmed to reach the vertically attached *A. japonicus*. *P. ortmanni* were not fed for a week before the experiments. For prey preparation, 100 *A. japonicus* juveniles were soaked in L-menthol solution for 20–30 min to induce anesthesia and estimate the standard body length (SBL), a widely applied method for accurately measuring sea cucumbers [33, 41]. After the measurements, juveniles were separately returned to the aquarium and carefully observed for several days to check their condition before the experiments. Ten randomly selected juveniles were gently placed in each aquarium as prey. Throughout all trials, a control aquarium without *P. ortmanni* was maintained to confirm that any observed mortality in *A. japonicus* was exclusively attributable to predation rather than alternative factors such as water quality, initial health status, or other variables. The SBL and wet weight of the tested *A. japonicus* were 12.34 ± 1.27 mm (mean \pm SD), ranging from 10.17 to 15.84 mm, and 0.03 ± 0.01 g, ranging from 0.01 to 0.07 g, respectively. The numbers of *A. japonicus* and predator–prey behavior were observed and noted at 1, 3, 6, and 12 h (the end of a light cycle) and 24 h (the end of a dark cycle) after the start of the experiments [33]. The experiments were repeated three times, and the studied *P. ortmanni* were immediately preserved in 70% ethanol to check their sex and measure their body size indices of SL, shield width (SW) at the widest part, right and left cheliped propodus length (CPL), wet weight, and shell characteristics, such as aperture length (AL) and aperture width (AW) (Figure 3a–c and Table 1). All measurements were taken with a digital vernier caliper to the nearest 0.01 mm. The *A. japonicus* juveniles used in this study were obtained from Hokkaido Aquaculture Promotion

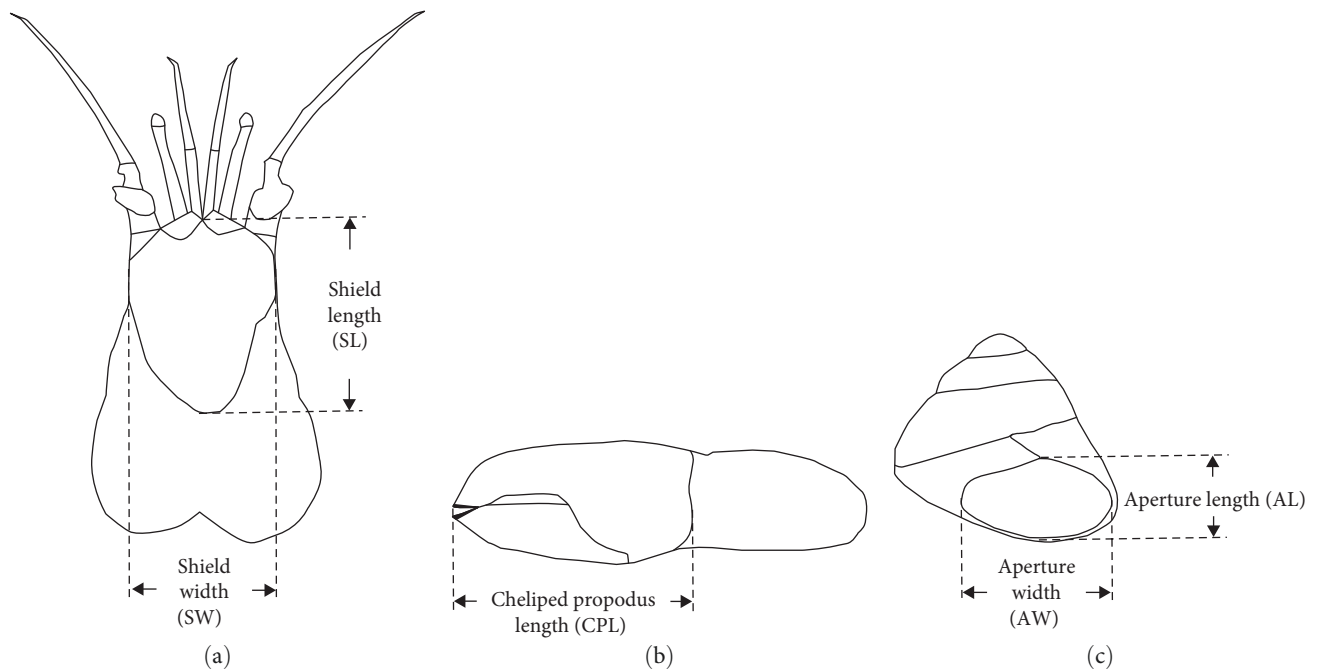


FIGURE 3: Shield length and width at the widest part (a), cheliped propodus length (b), and shell aperture length and aperture width (c) measured as body size and shell characteristics.

TABLE 1: Measured shield length (SL) and shield width (SW) at the widest part, the average of the right and left cheliped propodus length (CPL), shell aperture length (AL) and aperture width (AW), and wet weight of *P. ortmanni* ($n = 9$, two males and seven females) tested in the laboratory experiments.

Sample ID	Sex	Wet weight (g)	Shield length (mm)	Shield width (mm)	Average cheliped propodus length (mm)	Aperture length (mm)	Aperture width (mm)
1	F	0.516	6.56	5.52	4.26 ± 0.06	14.06	8.5
2	F	0.732	7.19	5.63	5.23 ± 0.09	14.59	7.34
3	F	0.398	5.84	4.47	4.06 ± 0.01	22.18	10.11
4	F	0.494	6.53	5.14	4.97 ± 0.03	13.59	11.10
5	F	0.645	6.38	5.66	5.05 ± 0.11	12.67	10.32
6	F	0.412	6.25	4.99	4.46 ± 0.05	14.29	8.35
7	F	0.560	6.85	6.10	5.41 ± 0.05	23.77	12.31
8	M	0.582	7.34	5.98	5.88 ± 0.07	24.69	11.60
9	M	0.899	7.92	6.28	6.67 ± 0.02	14.51	12.41

Corporation. By using Welch's *t*-tests, the differences in mortality rates between day and night, as well as sex in the predation experiments, were assessed. Correlation analyses (Pearson's correlation coefficient) were used to determine whether the mortality rate and each of the body size variables (SL, SW, CPL, and wet weight) and shell characteristic variables (AL and AW) were significantly correlated. Friedman tests were also applied to examine differences in the number of killed sea cucumbers in the three trials. All statistical analyses were performed using OriginPro 2020b (OriginLab Corporation, USA) and StatView statistical software (version 5.0).

2.3. Ethical Statement. The present study was performed in compliance with the relevant laws, institutional guidelines, and ethical guidelines for animal experiments by the Science Council of Japan and approved by the institutional ethics review board.

3. Results

3.1. Population Dynamics and Calcareous Ossicles in the Stomach Contents. The number of individuals, sex ratio, and SL frequency distributions of *P. ortmanni* during the sampling period are shown in Figure 4a–c. Although the population density of *P. ortmanni* showed similar values in June and September 2018, their population density increased to 59 individuals (c.a. 524 individuals per m^3) consisting of 34 males and 25 females in December, approximately threefold the population density in June and September. Both SL and wet weight also increased from June (6.02 ± 1.08 mm [mean \pm SD] and 0.71 ± 0.32 g) to December (7.41 ± 1.62 mm and 1.34 ± 0.97 g), and both values in December were found to be significantly higher than those in June and September ($p < 0.05$). A representative picture of *A. japonicus* ossicles found in stomach contents and

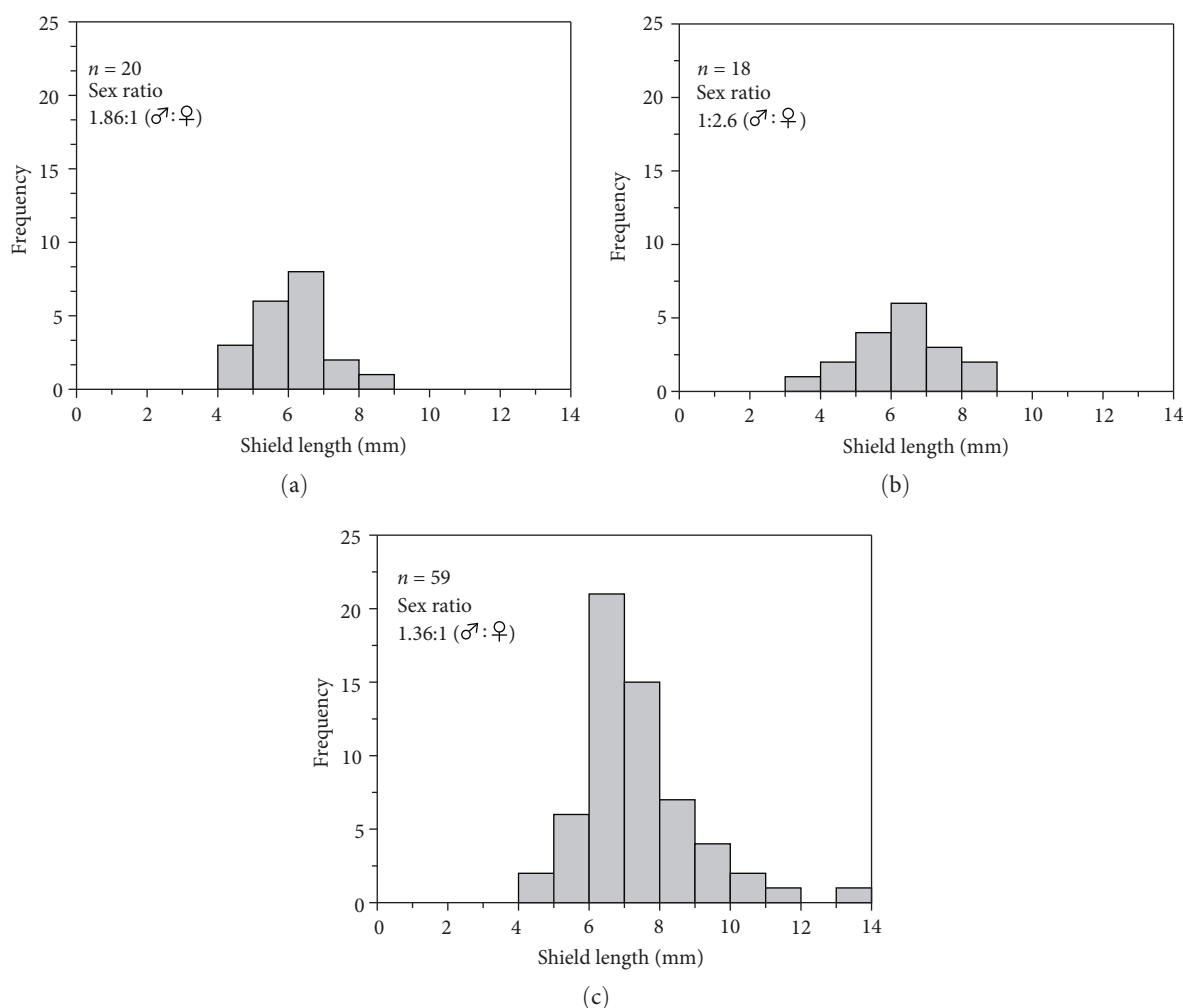


FIGURE 4: The number of individuals, sex ratio, and shield length (SL) frequency distributions of *P. ortmanni* in June (a), September (b), and December (c) 2018.

the ratio of ossicle-detected male-to-female *P. ortmanni* collected in December 2018 are shown in Figure 5a,b. Ossicles were detected in the stomach contents at a rate of 28.8% (18.6% males and 10.2% females). There were no statistically significant differences in ossicle detection rates between male and female hermit crabs ($p > 0.05$). The averages of the SL and wet weight of the ossicle-detected and not-detected groups were 7.85 ± 1.84 mm (mean \pm SD) ranging from 5.80 to 13.83 mm, 7.23 ± 1.48 mm ranging from 4.27 to 11.76 mm, 1.69 ± 1.31 g ranging from 0.49 to 6.11 g, and 1.20 ± 0.75 g ranging from 0.17 to 4.52 g, respectively. Although the average SL and wet weight were high in the ossicle-detected groups of both males and females, a statistically significant difference was observed only when comparing the average SL among female hermit crabs ($p < 0.01$) (Figure 6).

3.2. Estimating the Mortality Rate and Predator–Prey Behavior. The mortality of juvenile *A. japonicus* was observed in eight of nine aquariums regardless of the size or sex of the tested *P. ortmanni*, and the estimated average mortality rate was 2.5 ± 2.4 individuals day^{-1} (mean \pm SD) (Figure 7a). *P. ortmanni* preying on *A. japonicus* is also available as a video in

the Supporting Information (Video S1). Although the male *P. ortmanni* with the longest AL (24.69 mm) was capable of killing a maximum of nine juvenile *A. japonicus* within the first 12 h (Figure 7b,c), no statistically significant differences in mortality rates between the sexes were observed in these experiments. Although the studied hermit crabs foraged for food both during the day and night, the number of *A. japonicus* consumed during the day was significantly higher ($p < 0.001$). The mortality rate did not correlate significantly with any body size variables (SL, SW, CPL, or wet weight) or shell characteristic variables (AL or AW). On average, 3% to 5% of *A. japonicus* survived despite being attacked and injured in every study (Figure 7d), and noteworthy predation avoidance behavior of *A. japonicus* directly adhering to the shell of *P. ortmanni* was consistently observed in these experiments (Figure 7e). After adhering to the shell, most individuals remained on the shells until the end of the experiment.

4. Discussion

Most hermit crabs are believed to feed on detritus and carrion and are classified as “omnivorous detritivores” [42, 43].

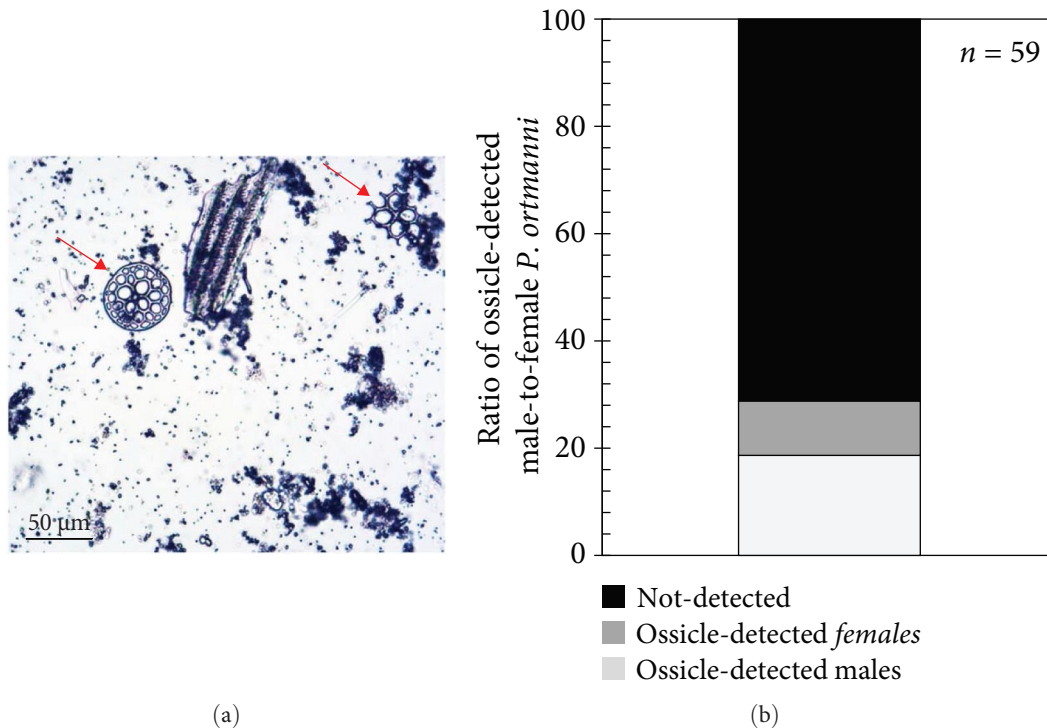


FIGURE 5: Representative picture of *A. japonicus* ossicles found in the stomach content of *P. ortmanni* (a) and the ratio of ossicle-detected male to female ($n = 59$) collected in December 2018 (b).

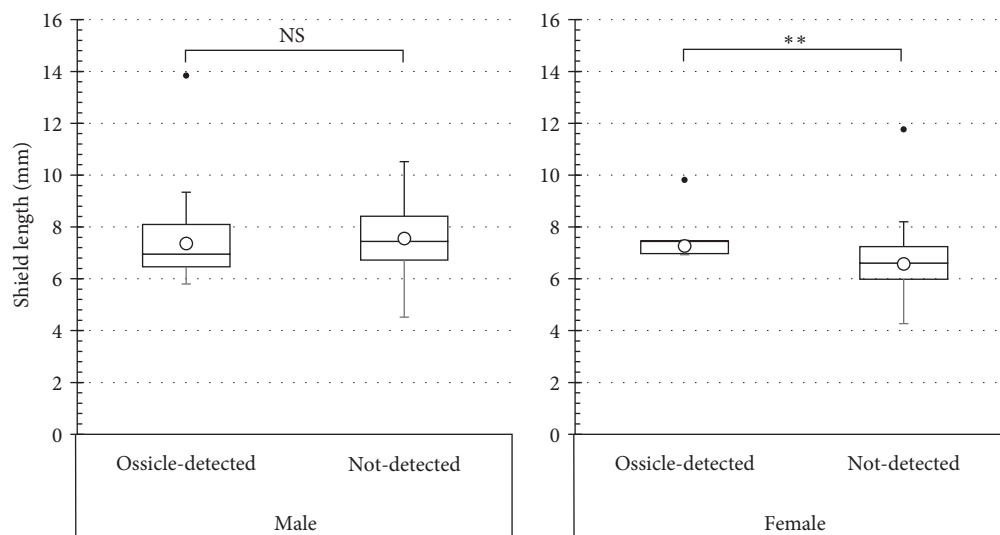


FIGURE 6: Boxplot of shield length (SL) between the ossicle-detected and not-detected groups in male and female *P. ortmanni* collected from the field in December 2018. Asterisk (**) denotes statistical significance ($p < 0.01$).

Still, some studies have demonstrated that living organisms form a substantial part of their diet [44], particularly hermit crabs belonging to the genera *Dardanus* and *Paguristes* [42, 45, 46]. In the present study, the hermit crab *Paguristes ortmanni*, known to overlap widely with the habitat of *A. japonicus* [37, 47–49], was identified as a predator of *A. japonicus* juveniles, as evidenced by the frequent detection of *A. japonicus* ossicles in their stomach contents at levels similar to those in the predatory crab *P. ferox* [33], along with the observed

predatory behavior. Although the male *P. ortmanni* (ID 8) predated nine juveniles within the first 12 h in one of the experiments, the estimated average mortality rate was about one-third that of *P. ferox*, which is considered to be the most powerful predator of juvenile *A. japonicus* [33]. However, the average mortality rate was higher than that of the sea star *Asterina pectinifera*, which was previously considered the principal predator of *A. japonicus* juveniles [19]. While several other sympatrically occurring decapod crustaceans, such

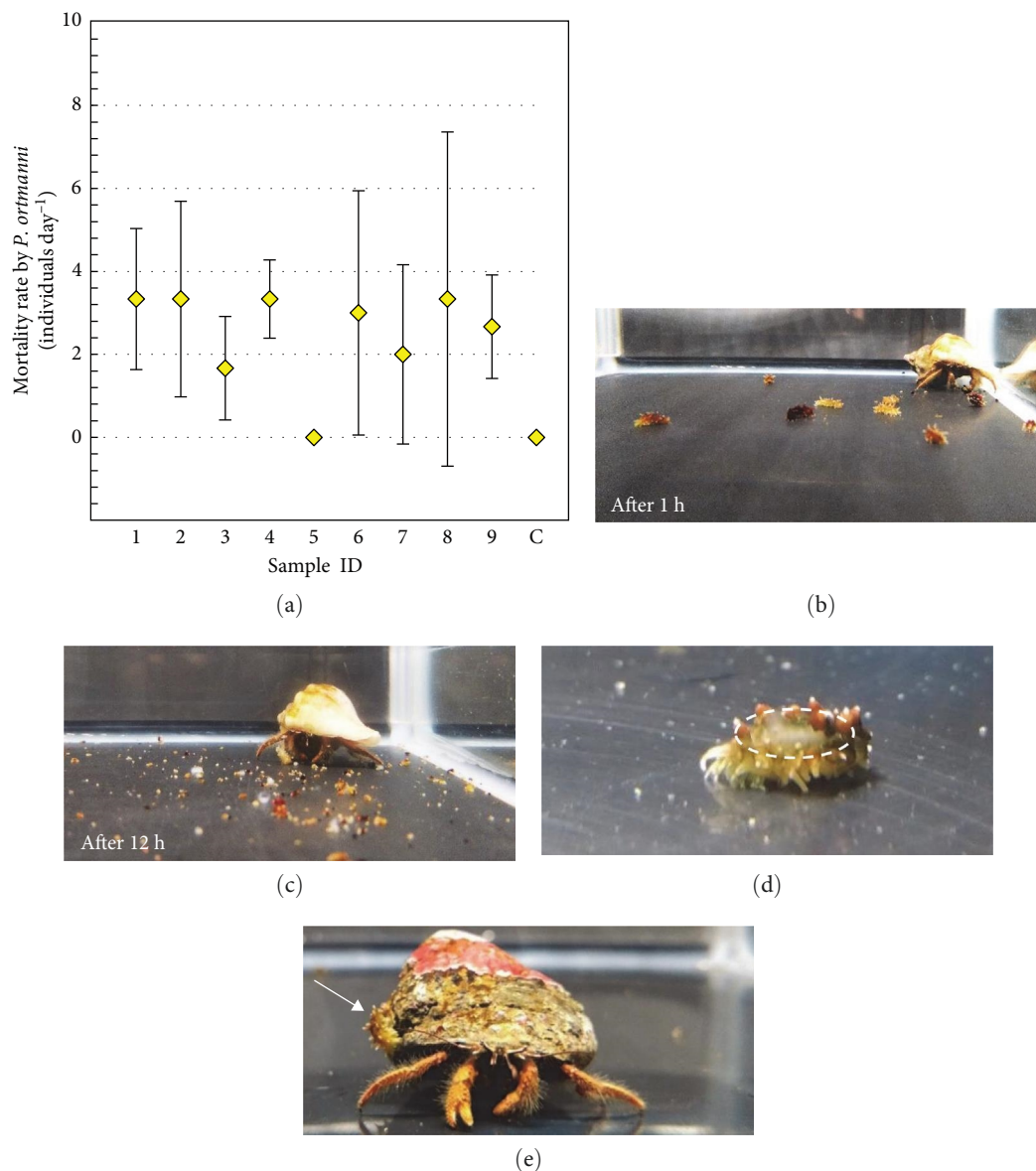


FIGURE 7: Estimated mortality rates in the laboratory predation experiments (a). An aquarium with no *P. ortmanni* was shown as the control (C). A picture of *P. ortmanni* (Sample ID: 8) with living and/or killed *A. japonicus* after 1 h (b) and 12 h (c), the surface body wall of *A. japonicus* ripped by *P. ortmanni* (d), and *A. japonicus* adhering to the shell of *P. ortmanni* (e).

as *P. ferox*, *Pagurus proximus*, and *Pagurus middendorffii*, were observed in the same area, the density of *P. ortmanni* reached approximately 524 individuals per m³, almost twice of the second most dominant crab *P. ferox*, in December 2018. In Hokkaido, *A. japonicus* juveniles are reared to approximately 10–30 mm at a land-based facility and then released to the wild from fall to winter [50, 51], which coincides with the period of population increase in the predatory hermit crab shown in this study.

Throughout the study, *A. japonicus* exhibited two types of common antipredatory behaviors, namely, muscle contraction and evisceration, and one unique and novel predation avoidance behavior of directly adhering to the shell of *P. ortmanni* (Figure 7e). *P. ortmanni* often struggled to feed, shifting prey from one hand to the other during feeding,

especially for larger juveniles, taking much longer than *P. ferox* [33]. Consequently, 3% to 5% of *A. japonicus* survived despite being attacked and injured in every trial. Releasing size is one of the key factors in improving the survival of released juveniles for a range of invertebrates, including sea cucumber [52], abalone, clam, topshell [28], and crab [53], and is often explained by a higher tolerance to environmental stressors (water temperature and salinity) and predator-induced mortality [52, 54]. It is not certain whether the predation avoidance behavior observed in this study occurs in a natural environment, but the escape behavior of *A. japonicus* that takes advantage of the blind spot of hermit crabs accompanied by detachment [23, 24] may be very effective in escaping predation. The feeding behavior of decapod crustaceans, such as feeding rate and prey handling,

is known to be influenced not only by the size of the prey but also by the size and sex of the predator [33, 55–57]. In this study, more than half of the measured SL of the not-detected group (females) were smaller than that of the smallest individual in the ossicle-detected group, and the average SL of the ossicle-detected group in females was significantly greater than that of the not-detected group. These results indicate an elevated predation risk of *A. japonicus* juveniles by larger female *P. ortmanni*. However, in the laboratory experiments, there were no significant differences in mortality rates between sexes, and the mortality rate showed no significant correlation with any body size or shell characteristic variables. The small sample size and imbalance in the male-to-female ratio of the hermit crabs used in the experiments may account for these results. Numerous studies have also reported the influence of the developmental stage, feeding selectivity, and environmental conditions of decapod crustaceans on feeding behaviors [58, 59]. Consequently, future research from these perspectives may provide a more comprehensive understanding of relationships with predatory organisms that contribute to enhancing the population of commercially important endangered *A. japonicus*.

5. Conclusions

This study presents the first evidence that the sympatrically occurring hermit crab *P. ortmanni* is a predator of *A. japonicus* juveniles, emphasizing the necessity of comprehending their distribution and occurrence in regions where hatchery-produced juveniles are released. Furthermore, this research underscores the significance of implementing appropriate measures to ensure successful wild release, such as avoiding areas and periods with high predatory hermit crab density, utilizing physical enclosures to prevent predator invasion, and selecting an appropriate release size to mitigate predation risk.

Data Availability Statement

The original contributions presented in the study are included in the article/supporting information; further inquiries can be directed to the corresponding author.

Conflicts of Interest

Yuji Anaguchi was employed by Ocean Construction Co., Ltd. The remaining authors declare that the research was conducted without any commercial or financial relationships that could be construed as a potential conflicts of interest.

Author Contributions

Nobuharu Inaba: conceptualization, investigation, analysis, writing—original draft, supervision. **Takuma Matsumoto:** investigation, writing—review and editing. **Yuji Anaguchi:** investigation, writing—review and editing. **Kohei Matsuno:** resources, writing—review, editing, supervision.

Funding

This research was conducted by a management expense grant from Public Works Research Institute, Japan, and did not receive any external funding.

Acknowledgments

We express sincere gratitude to the members of the Fisheries Engineering Research Team, Civil Engineering Research Institute for Cold Region, Japan, for administrative support, especially Sawako Shirai, a technical assistant. We would also like to show our great appreciation to Professor Satoshi Wada from Hokkaido University for his essential advice about hermit crab identification. Our sincere appreciation goes to Dr. Atsushi Yamaguchi from Hokkaido University for providing laboratory space during the investigation.

Supporting Information

Additional supporting information can be found online in the Supporting Information section. (*Supporting Information*) Video S1: Video of hermit crab *P. ortmanni* preying on juvenile sea cucumber *A. japonicus*.

References

- [1] A. Mercier, A. Gebruk, A. Kremenetskaia, and J. F. Hamel, “An Overview of Taxonomic and Morphological Diversity in Sea Cucumbers (Holothuroidea: Echinodermata),” in *The World of Sea Cucumbers*, Academic Press, eds. A. Mercier, J. F. Hamel, A. D. Suhrbier, and C. M. Pearce, 3, 2024): 15.
- [2] H. Zhao, “Taxonomy and Identification,” in *The Sea Cucumber *Apostichopus japonicus*: History, Biology and Aquaculture*, eds. H. S. Yang, J. F. Hamel, and A. Mercier, 37, (Academic Press, Amsterdam, Netherlands, 2015): 52.
- [3] V. Toral-Granda, A. Lovatelli, and M. Vasconcellos, “Sea Cucumbers. A Global Review of Fisheries and Trade,” in *FAO Fisheries and Aquaculture Technical Paper (FAO, Rome)*, eds. V. Toral-Granda, A. Lovatelli, and M. Vasconcellos, 2008): 317.
- [4] H. Eriksson, H. Österblom, B. Crona, et al., “Contagious Exploitation of Marine Resources,” *Frontiers in Ecology and the Environment* 13 (2015), 435–440.
- [5] S. Purcell, B. A. Polidoro, J. F. Hamel, R. U. Gamboa, and A. Mercier, “The Cost of Being Valuable: Predictors of Extinction Risk in Marine Invertebrates Exploited as Luxury Seafood. Proc. R.,” *Soc B* 281, 20133296 (2014).
- [6] P. S. Choo, “Population Status, Fisheries and Trade of Sea Cucumbers in Asia, A Global Review of Fisheries and Trade,” in *FAO Fisheries and Aquaculture Technical Paper*, eds. V. Toral-Granda, A. Lovatelli, M. Eds Vasconcellos, and Sea Cucumbers, no. 516, (Rome, Italy, 2008): 81–118.
- [7] J. F. Hamel and A. Mercier, “*Apostichopus japonicus*. The IUCN Red List of Threatened Species,” (2013): e.T180424A1629389.
- [8] N. P. Brown and S. D. Eddy, *Echinoderm Aquaculture* (Wiley Blackwell, Hoboken, USA: 384, 2015).
- [9] J. Akamine, “Challenging “boom and Bust” Market Pressures: Development of Self-Managed Sea Cucumber Conservation in Rishiri Island, Hokkaido, Japan,” *Biosphere Conservation: for Nature, Wildlife, and Humans* 9 (2009): 1–12.
- [10] J. Akamine, “*Apostichopus japonicus*: Fisheries, Trade, and Foodways in Japan,” in *Developments in Aquaculture and*

- Fisheries Science*, eds. H. Yang, J. F. Hamel, and A. Mercier, 39, (Elsevier, 2015): 399–421.
- [11] MAFF, “Poaching Shall never Be Forgiven: Fisheries. The Ministry of Agriculture, Forestry and Fisheries,” (2021).
 - [12] X. Yuan, S. Shao, S. Dupont, L. Meng, Y. Liu, and L. Wang, “Impact of CO₂ Driven Acidification on the Development of the Sea Cucumber *Apostichopus japonicus* (Selenka) (Echinodermata: Holothuroidea),” *Marine Pollution Bulletin* 95, no. 1 (2015): 195–199.
 - [13] X. Yuan and X. Xie, “Sea Cucumbers Under Ocean Acidification and Warming,” in *The World of Sea Cucumbers: Challenges Advances, and Innovations*, eds. A. Mercier, J. F. Hamel, A. D. Suhrbier, and C. M. Pearce, (Academic Press, Amsterdam, Netherlands, 2024): 483–492.
 - [14] T. Qiu, T. Zhang, J. F. Hamel, and A. Mercier, “Development, Settlement, and Post-Settlement Growth,” in *The Sea Cucumber *Apostichopus japonicus*: History, Biology and Aquaculture*, eds. H. S. Yang, J. F. Hamel, and A. Mercier, (Academic Press, Amsterdam, Netherlands, 2015): 111–131.
 - [15] S. Goshima, “Ecology of Japanese Sea Cucumber,” in *Namako Gyogyo to sSono Kanri: Shigen, Seisan, Shijo, Koseisha-kKoseikaku*, eds. M. Hirota and M. Machiguchi, 47, (Tokyo, 2014): 71.
 - [16] S. Uthicke, D. Welch, and J. A. H. Benzie, “Slow Growth and Lack of Recovery in Overfished Holothurians on the Great Barrier Reef: Evidence from DNA Fingerprints and Repeated Large-Scale Surveys,” *Conservation Biology* 18 (2004): 1395–1404.
 - [17] Hokkaido Government, “Hokkaido Fisheries Today. Department of Fisheries and Forestry,” 121 (2022).
 - [18] P. Francour, “Predation on Holothurians: A Literature Review,” *Invertebrate Biology* 116 (1997): 52–60.
 - [19] Z. Yu, H. Yang, and J. F. Hamel, “Larval, Juvenile, and Adult Predators,” in *The Sea Cucumber *Apostichopus japonicus*: History, Biology and Aquaculture*, eds. H. S. Yang, J. F. Hamel, and A. Mercier, (Academic Press, Amsterdam, Netherlands, 2015): 243–256.
 - [20] Y.-C. Zhao, C.-H. Xue, T.-T. Zhang, and Y.-M. Wang, “Saponins from Sea Cucumber and Their Biological Activities,” *Journal of Agricultural and Food Chemistry* 66, no. 28 (2018): 7222–7237.
 - [21] C. Legault and J. H. Himmelman, “Relation Between Escape Behavior of Benthic Marine Invertebrates and the Risk of Predation,” *Journal of Experimental Marine Biology and Ecology* 170, no. 1 (1993): 55–74.
 - [22] J. J. So, J. F. Hamel, and A. Mercier, “Habitat Utilisation, Growth and Predation of *Cucumaria frondosa*: Implications for an Emerging Sea Cucumber Fishery,” *Fisheries Management and Ecology* 17, no. 6 (2010): 473–484.
 - [23] B. Morton, “Aspects of Predation by *Tonna Zonatum* (Prosobranchia: Tonnoidea) Feeding on Holothurians in Hong-Kong,” *Journal of Molluscan Studies* 57, no. 1 (1991): 11–19.
 - [24] J. F. Hamel, J. Sun, B. Gianasi, et al., “Active Buoyancy Adjustment Increases Dispersal Potential in Benthic Marine Animals,” *Journal of Animal Ecology* 88, no. 6 (2019): 820–832.
 - [25] R. H. Emson and I. C. Wilkie, “Fission and Autotomy in Echinoderms,” *Oceanography and Marine Biology: An Annual Review* 18 (1980): 155–250.
 - [26] L. A. Gosselin and P. Y. Qian, “Juvenile Mortality in Benthic Marine Invertebrates,” *Marine Ecology Progress Series* 146 (1997): 265–282.
 - [27] H. L. Hunt and R. E. Scheibling, “Role of Early Post-Settlement Mortality in Recruitment of Benthic Marine Invertebrates,” *Marine Ecology Progress Series* 155 (1997): 269–301.
 - [28] J. D. Bell, J. L. Munro, W. J. Nash, et al., “Restocking and Stock Enhancement of Marine Invertebrate Fisheries,” in *Advances in Marine Biology*, 49, (Academic Press, London, UK, 2005): 392.
 - [29] S. K. Dance, I. Lane, and J. D. Bell, “Variation in Short-Term Survival of Cultured Sandfish (*Holothuria scabra*) Released in Mangrove-Seagrass and Coral Reef Flat Habitats in Solomon Islands,” *Aquaculture* 220, no. 1–4 (2003): 495–505.
 - [30] T. Lavitra, R. Rasolofonirina, M. Jangoux, and I. Eeckhaut, “Problems Related to the Farming of *Holothuria scabra* (Jaeger, 1833),” *SPC Beche-de-Mer Information* 29 (2009): 20–30.
 - [31] T. Lavitra, G. Tsiresy, R. Rasolofonirina, and I. Eeckhaut, “Effect of Nurseries and Size of Released *Holothuria scabra* Juveniles on Their Survival and Growth,” *SPC Beche-de-Mer Information* 35 (2015): 37–41.
 - [32] O. J. C. Caasi, L. A. Gosselin, and M. A. Juinio-Meñez, “Size-Dependent Predation on Juvenile Sandfish, *Holothuria scabra* by Seagrass-Associated Crabs,” *Journal of the Marine Biological Association of the United Kingdom* 103 (2023): 1–13.
 - [33] N. Inaba, T. Matsumoto, H. Kawai, Y. Anaguchi, and K. Matsuno, “Predation of Juvenile Japanese Sea Cucumber *Apostichopus japonicus* by Kelp Crab *Puggettia Ferox*,” *Frontiers in Marine Science* 8 (2021): 684989.
 - [34] Y. Yamana, Y. Furukawa, S. Kashio, and S. Goshima, “Environmental Characteristics of the Habitats of Juvenile Sea Cucumber *Apostichopus japonicus* Around Hokkaido. -Several Inferences From Southern Hokkaido,” *Aquaculture Sci* 62, no. 2 (2014): 163–181.
 - [35] S. Miyake, “The Crustacean Anomura of Sagami Bay,” *Biological Laboratory, Imperial Household (Tokyo)* 1 (1978).
 - [36] T. Komai, “A Review of the North-Western Pacific Species of the Genus *Paguristes* (Decapoda: Anomura: Diogenidae), I. Five Species Initially Reported by Ortmann, 1892 From Japan,” *Journal of Natural History* 35, no. 3 (2001): 357–428.
 - [37] V. V. Petryashov and E. S. Kornienko, “*Paguristes Ortmanni*, Miyake, 1978 (Decapoda: Anomura) - A New Genus and Species of Decapods for the Russia Region,” *Russian Journal of Marine Biology* 32, no. 2 (2006): 120–122.
 - [38] P. A. McLaughlin, T. Komai, R. Lemaitre, and D. L. Rahayu, “Annotated Checklist of Anomuran Decapod Crustaceans of the World (Exclusive of the Kiwaoidea and Families Chirostylidae and Galatheidae of the Galatheoidea) Part I,” *Lithodoidea, Lomisoidea and Paguroidea. Raffles Bull. Zool Supplement* 23 (2010): 5–107.
 - [39] F. Gao and H. Yang, “Anatomy, the Sea Cucumber *Apostichopus japonicus*: History, Biology and Aquaculture,” vol. 53, (Academic Press, Amsterdam, Netherlands, 2015): 76.
 - [40] J. W. Tukey, *Exploratory Data Analysis*, 18 (Addison-Wesley Publishing Company, (2nd edition) edition, 1977): 688.
 - [41] Y. Yamana, S. Goshima, T. Hamano, T. Yusa, Y. Furukawa, and N. Yoshida, “Formulae to Estimate Standard Body Length for Regional Forms of the Sea Cucumber *Apostichopus japonicus* in Japan,” *Nippon Suisan Gakkaishi* 77, no. 6 (2011): 989–998.
 - [42] B. A. Hazlett, “The Behavioral Ecology of Hermit Crabs,” *Annual Review of Ecology and Systematics* 12, no. 1 (1981): 1–22.

- [43] I. Lancaster, "Pagurus bernhardus (L.) - an Introduction to the Natural History of Hermit Crabs," *Field Studies* 7 (1988): 189–238.
- [44] I. Volvenko, "Nutrition and Feeding Behavior of Hermit Crabs," *Russian Journal of Marine Biology* 20 (1994): 307–313.
- [45] P. J. Schembri, "Feeding Behavior of Fifteen Species of Hermit Crabs (Crustacea: Decapoda: Anomura) From the Otago Region, Southeastern New Zealand," *Journal of Natural History* 16, no. 6 (2007): 859–878.
- [46] C. F. MacKay, C. B. Untiedt, and L. Hein, "Local Habitat Drivers of Macrobenthos in the Northern, Central and Southern KwaZulu-Natal Bight, South Africa," *African Journal of Marine Science* 38, no. sup1 (2016): S105–S121.
- [47] H. S. Kim, "A Checklist of the Anomura and Brachyura (Crustacea, Decapoda) of Korea," *Journal of Seoul National University Biological Sciences* 21 (1970): 1–29.
- [48] A. Asakura, "Shallow Water Hermit Crabs of the Families Pylochelidae, Diogenidae and Paguridae (Crustacea: Decapoda: Anomura) From the Sea of Japan, With a Description of a New Species of *Diogenes*," *Bulletin of the Toyama Science Museum* 29 (2006): 23–103.
- [49] J. Liu, "Spatial Distribution, Population Structures, Management, and Conservation," in *The Sea Cucumber Apostichopus japonicus: History, Biology and Aquaculture*, eds. H. S. Yang, J. F. Hamel, and A. Mercier, 77, (Academic Press, Amsterdam, Netherlands, 2015): 86.
- [50] Y. Sakai, "Mass Production of Artificial Seed of the Japanese Common Sea Cucumber (*Apostichopus japonicus*) in Hokkaido, Japan," *Bulletin of Japan Fisheries Research and Education Agency* 40 (2015): 129–134.
- [51] Q. Han, J. K. Keesing, and D. Liu, "A Review of Sea Cucumber Aquaculture, Ranching, and Stock Enhancement in China," *Reviews in Fisheries Science & Aquaculture* 24, no. 4 (2016): 326–341.
- [52] S. Purcell and M. Simutoga, "Spatio-Temporal and Size-Dependent Variation in the Success of Releasing Cultured Sea Cucumbers in the Wild," *Reviews in Fisheries Science* 16, no. 1–3 (2008): 204–214.
- [53] E. G. Johnson, A. H. Hines, M. A. Kramer, and A. C. Young, "Importance of Season and Size of Release to Stocking Success for the Blue Crab in Chesapeake Bay," *Reviews in Fisheries Science* 16, no. 1–3 (2008): 243–253.
- [54] H. Hatanaka, H. Uwaoku, and T. Yasuda, "Experimental Studies on the Predation of Juvenile Sea Cucumber, *Stichopus Japonicus* by Seastar, *Asterina pectinifera*," *Suisanzoshoku* 42 (1994): 563–566.
- [55] S. Y. Lee and R. Seed, "Ecological Implications of Cheliped Size in Crabs: Some Data From *Carcinus maenas* and *Liocarcinus holsatus*," *Marine Ecology Progress Series* 84 (1992): 151–160.
- [56] J. Freire, M. P. Sampedro, and E. González-Gurriarán, "Influence Morphometry and Biomechanics on Diet Selection in Three Portunid Crabs," *Marine Ecology Progress Series* 137 (1996): 111–121.
- [57] J. M. Kolts, J. R. Lovvorn, C. A. North, J. M. Grebmeier, and L. W. Cooper, "Effects of Body Size, Gender, and Prey Availability on Diets of Snow Crabs in the Northern Bering Sea," *Marine Ecology Progress Series* 483 (2013): 209–220.
- [58] S. S. L. Lim, A. Y. P. Yong, and J. H. Christy, "Ontogenetic Changes in Diet and Related Morphological Adaptations in *Ocypode gaudichaudii*," *Invertebrate Biology* 135, no. 2 (2016): 117–126.
- [59] P. Briones-Fourzán and M. E. Hendrickx, "Ecology and Diversity of Marine Decapod Crustaceans," *Diversity* 14, no. 8 (2022): 614.